Powerful background reduction in fluorescent tissue stains with an improved proximity-based technology for detection of protein-protein interactions

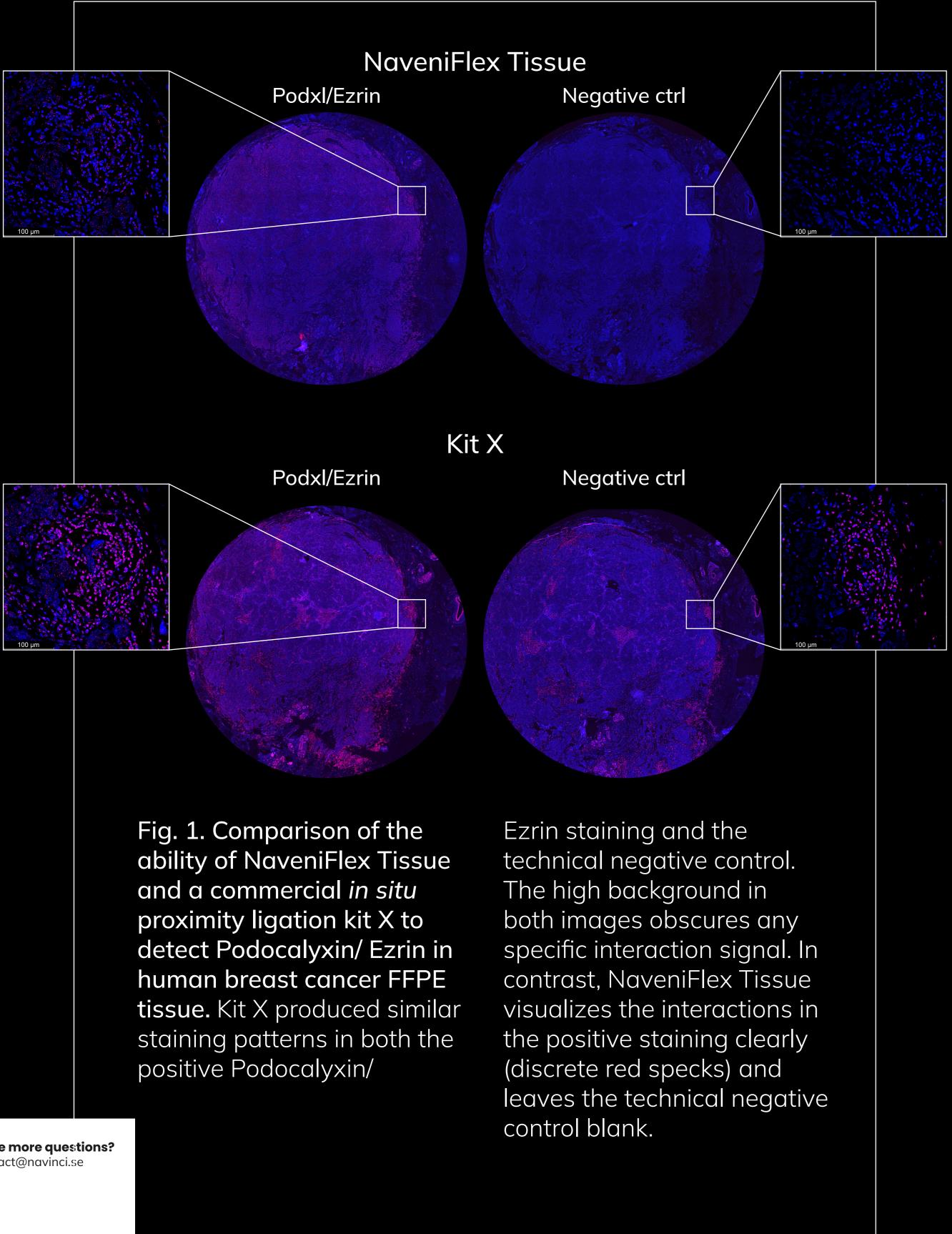
Authors

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Introduction

Immunofluorescent staining of tissues via in situ proximity ligation assays is a well-established tool for highly sensitive detection of protein-protein interactions (PPI), post-translational modifications (PTM), and their localization. Standard immunofluorescence (IF) and in situ proximity ligation assays suffer from

background signal originating from unspecific binding of fluorescently labeled detection reagents to certain types of cells (cell-specific background). As a result, it can be difficult to distinguish true biologically relevant signal from background signal. To address this problem, we developed

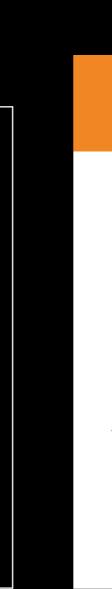


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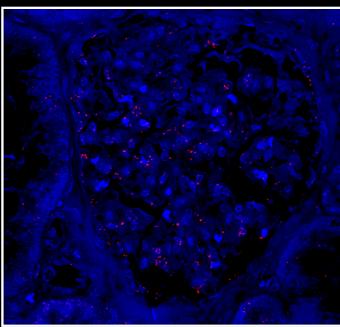


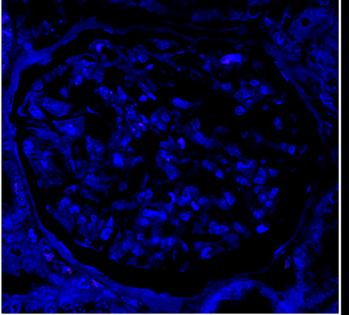
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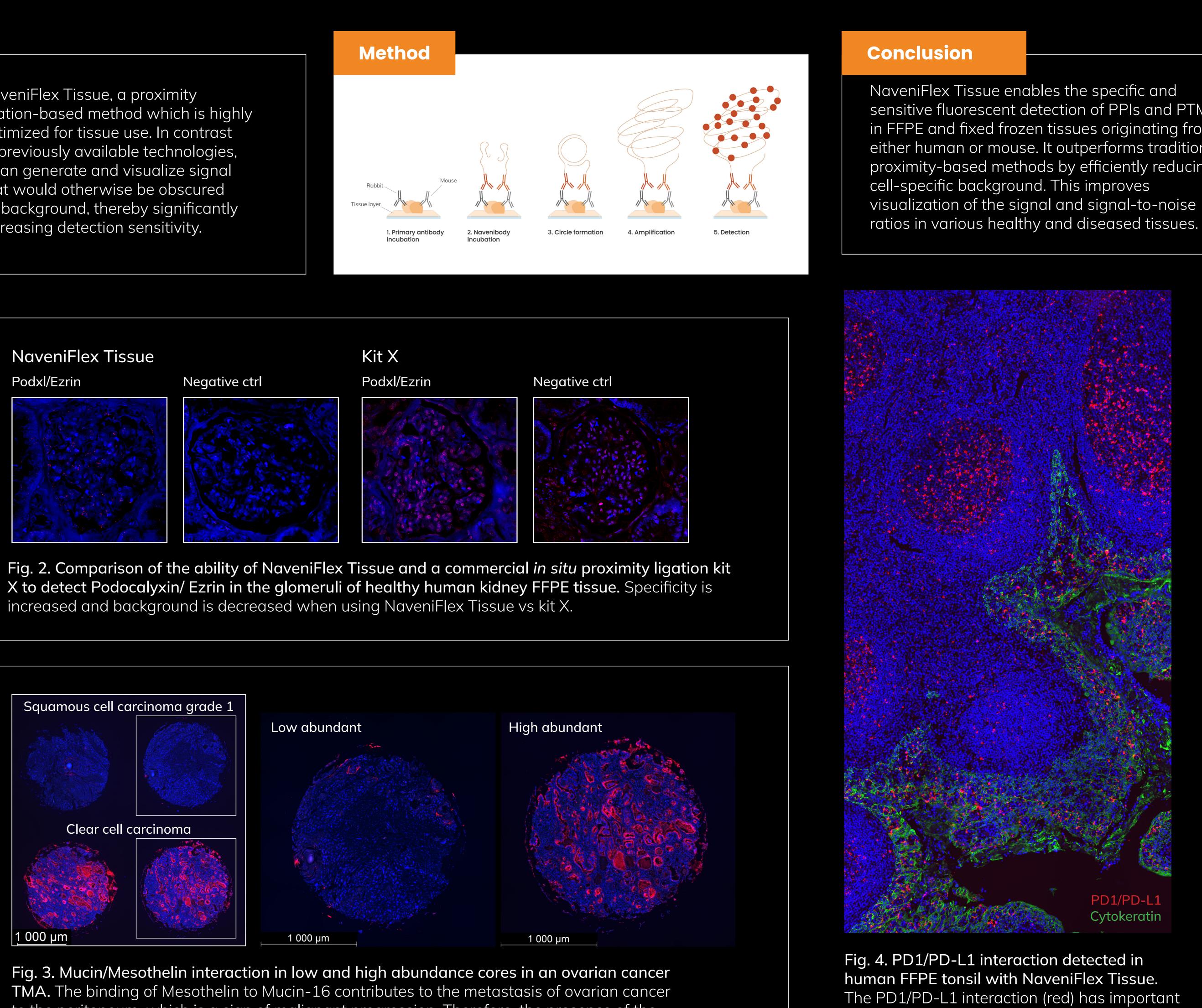
NaveniFlex Tissue, a proximity ligation-based method which is highly optimized for tissue use. In contrast to previously available technologies, it can generate and visualize signal that would otherwise be obscured by background, thereby significantly increasing detection sensitivity.



Podxl/Ezrin







to the peritoneum, which is a sign of malignant progression. Therefore, the presence of the interaction may have prognostic significance in determining patient relapse-free survival. NaveniFlex Tissue successfully stains both sparse (grade 1 squamous cell carcinoma) and highly abundant (clear cell carcinoma) interactions (see zoom-ins).

The PD1/PD-L1 interaction (red) has important implications in immunooncology and has been notoriously difficult to detect with older proximity ligation methods. NaveniFlex Tissue, however, detects this prognostically important interaction. IF cytokeratin co-staining in green.



sensitive fluorescent detection of PPIs and PTMs in FFPE and fixed frozen tissues originating from either human or mouse. It outperforms traditional proximity-based methods by efficiently reducing